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Tetrahedron Letters

Tetrahedron Letters 48 (2007) 4229–4233

# Total synthesis and morphogenesis-inducing activity of (±)-thallusin and its analogues

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> Received 19 March 2007; revised 9 April 2007; accepted 13 April 2007 Available online 20 April 2007

Abstract—Total synthesis of  $(\pm)$ -thallusin was achieved using Hg(OTf)<sub>2</sub>PhNMe<sub>2</sub>-induced olefin cyclization, and Suzuki coupling with a pyridylboronic acid derivative. Hg(OTf)<sub>2</sub> also acted as a catalyst to isomerize the double bond into the more thermodynamically stable isomer when treated in toluene. Synthetic  $(\pm)$ -thallusin as well as an analogue showed morphogenesis-inducing activity.  $© 2007 Elsevier Ltd. All rights reserved.$ 

Produced by a marine bacterium, thallusin is an algal morphogenesis inducer, that is, indispensable for the foliaceous morphology of macroalgae. Thallusin, which was isolated in 2005 after pursuing for more than 50 years, showed foliaceous morphology-inducing activity at the exceptionally low concentration of 1 ag/mL at the first stage. Structure 1 (Scheme 1) was established via a spectral study as well as via the single-crystal X-ray diffraction study of a thallusin derivative.<sup>[1](#page-3-0)</sup> In 2006, Snider and co-workers achieved the total synthesis of compound 1 from sclareol oxide 2. [2](#page-3-0) Because compound 1 did not show the foliaceous morphogenesis-inducing



Scheme 1.

activity, the absolute structure of thallusin was determined to be antipodal 3. Thallusin can be obtained in very limited amounts from cultivation. Thus, in the present study, the total syntheses of  $(\pm)$ -thallusin and analogues were undertaken to allow a detailed examination of thallusin's biological activity.

In 1983, we developed  $Hg(OTf)$  as a highly efficient olefin cyclization agent.[3](#page-3-0) In the present study, olefin cyclization in  $CH<sub>3</sub>NO<sub>2</sub>$  was the fundamental strategy to construct the carbon framework of 3. Due to its significant  $\pi$ -philicity, Hg(OTf)<sub>2</sub> in toluene also acts as a catalyst to isomerize the double bond into the thermodynamically more stable isomer.[4](#page-3-0) Suzuki coupling using a pyridylboronic acid derivative was employed for the introduction of an aromatic residue.<sup>[5](#page-3-0)</sup> Herein, we report a total synthesis of  $(\pm)$ -thallusin using homofarnesyl acetate (4, [Scheme 2\)](#page-1-0) as the starting material, and the morphogenesis-inducing activities of the synthetic  $(\pm)$ thallusin, its synthetic analogues, and some natural product derivatives.

Homofarnesyl acetate (4) was prepared in a total 91% yield from E,E-farnesol in four steps (a) bromination with  $PBr_3$ , (b) coupling of the Grignard reagent derived from chloromethyldimethylisopropoxysilane in the presence of CuI, (c) oxidative cleavage by the Tamao protocol using  $KH_2F$ ,  $H_2O_2$ , and  $Na_2CO_3$ ,<sup>[6](#page-3-0)</sup> and (d) acetylation with  $Ac_2O$  in pyridine. The reaction of 4

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<span id="page-1-0"></span>

#### Scheme 2.

with a complex of  $Hg(OTf)_2$  and N,N-dimethylaniline  $(1.2 \text{ equiv})$  was achieved in CH<sub>3</sub>NO<sub>2</sub> at  $-20 \text{ °C}$  for 3 h (Scheme 2). The crude extract was directly reacted with LAH in THF at  $0^{\circ}$ C for 1 h and the resulting alcohol was acetylated with acetic anhydride and pyridine to give rise to a mixture (double bond regio-isomers as well as stereoisomers at C-9) of acetates 5 in 94% yield. To reduce the number of isomers of 5, we first examined isomerization according to the Grieco and Nishizawa protocol using  $5 \text{ mol } \%$  of RhCl<sub>3</sub> $3H_2O$  in EtOH at 100 °C for 10 h.<sup>[7](#page-3-0)</sup> However, a complex mixture was formed. Then, we examined the reaction of 5 with catalytic amount of Hg(OTf)<sub>2</sub>, which has significant  $\pi$ -philic-ity.<sup>[4](#page-3-0)</sup> The original 4:3 mixture of  $\Delta^8$ -5 and  $\Delta^7$ -5 had converted into a 9:1 mixture on treatment with 5 mol % of  $Hg(OTf)<sub>2</sub>$  in toluene at room temperature

for 2 h. The 9:1 ratio seemed to be the final equilibrium composition because this ratio did not change after 24 h. The acetyl group of 6 was cleaved by the treatment with NaOMe in MeOH and the subsequent oxidation with TPAP afforded an aldehyde 7 in quantitative yield. The enolate prepared from  $8^8$  $8^8$  with LDA in THF was treated with aldehyde 7 to give alcohol 9 as a complicated diastereomeric mixture in 80% yield. LAH reduction of 9 in THF at  $0^{\circ}$ C afforded diol 10, also as a complex mixture of stereoisomers, in 70% yield. Treatment of 10 with triphosgene (2 equiv) in  $CH_2Cl_2$  in the presence of pyridine (5 equiv) at  $0^{\circ}$ C for 2 h afforded carbonate 11 in 76% yield. The *t*-butyl group was cleaved by using 20 equiv of TFA in CH<sub>2</sub>Cl<sub>2</sub> at  $0^{\circ}$ C for 2 h to give alcohol 12 in 73% yield. Reaction of 12 with 6 equiv of  $BF_3E_2O$  in  $CH_2Cl_2$  at 0 °C to room

temperature for 12 h selectively afforded the expected *trans*-cyclization product 13 selectively in  $52\%$  yield.<sup>[9](#page-3-0)</sup> Upon exposure of 13 to aqueous NaOH in dioxane at room temperature for 5 min, diol 14 was provided in 95% yield. The primary alcohol of 14 was selectively protected with a TBS group by reaction with TBSCl, DMAP, and Et<sub>3</sub>N in CH<sub>2</sub>Cl<sub>2</sub> at 0 °C for 4 h. Subsequent oxidation with Dess–Martin periodinate in  $CH_2Cl_2$  at  $0^{\circ}$ C to room temperature for 12 h afforded ketone 15 as a mixture of two stereoisomers in 69% yield. After the treatment of 15 with LDA at  $-78$  °C, the resulting lithium enolate was treated with Comins reagent 16 at  $-78$  °C to room temperature for 4 h, giving rise to the kinetically controlled enol triflate  $17$  in  $60\%$  yield.<sup>10</sup> The unstable enol triflate 17 was mixed with pyridylboronic acid  $18$ ,<sup>[11](#page-3-0)</sup> Pd(Ph<sub>3</sub>P)<sub>4</sub>, Ba(OH)<sub>2</sub>, and LiCl in aqueous dioxane and heated to 95  $\degree$ C for 2 h, giving rise to coupling product 19 in  $81\%$  yield.<sup>[5,12](#page-3-0)</sup> The TBS group of 19 was cleaved by treatment with HF–pyridine in THF at  $0^{\circ}$ C for 2 h to give the primary alcohol 20 in 98% yield. Compound 20 was subjected to Dess–Martin oxidation in  $\widehat{\mathrm{CH}_2Cl_2}$  at 0 °C to room temperature for 2 h, followed by  $NaClO<sub>2</sub>$  oxidation in the presence of  $NaH<sub>2</sub>PO<sub>4</sub>$ , and 2-methyl-2-butene in aqueous *t*-BuOH. Subsequent reaction with TMS diazomethane afforded the  $\alpha$ ,  $\beta$ -unsaturated ester 21 in 57% yield (total of three steps). The double bond of 20 had migrated to the conjugated position during these oxidation steps. Reaction of 21 with  $Pd(OAc)<sub>2</sub>$  dppf and NaOAc in MeOH under CO atmosphere  $(15 \text{ bar})$  at 135 °C for 10 h afforded trimethyl ester 22 in  $77\%$  yield.<sup>13</sup> The <sup>1</sup>H NMR spectrum of 22 was indistinguishable from that of the natural thallusin trimethyl ester. Finally, the methyl groups on 22 were cleaved by treatment with NaOD in  $D_2O$  at  $100 \degree C$  with NMR monitoring. After 2 h, trisodium tricarboxylates of  $(\pm)$ -thallusin  $[(\pm)$ -23] were obtained in quantitative yield.

During this synthetic program, several problems were encountered. Initially the alkylation of aldehyde 7 was performed with an anion derived from 24 (Scheme 3).[14](#page-3-0) After the acetylation, the ethoxyethyl protecting group was cleaved by methanolysis affording 25. Treatment of hydroxyacetate 25 with a variety of acids afforded a mixture of 26 and 27. Compound 27 was always the major product formed after acetyl group migration. Acetate 27 was further transformed into stable enol triflate 28 by methanolysis, Dess–Martin oxidation and treatment with LDA and then Comins' reagent 16.<sup>[10](#page-3-0)</sup> The structure of 28 was confirmed by a single-crystal X-ray diffraction study.[15](#page-3-0) Alternatively, the five-membered ring ether 28 was selectively prepared by reacting benzyl ether 24b with 7 via 27b. Suzuki coupling of 28 with 18 afforded 29 in poor yield probably due to the electron-deficient conjugated system. In contrast, the reaction of nonconjugated enol triflate 17 with 18 occurred readily, affording 19. Carbomethoxylation of 29 provided 30, and alkaline hydrolysis of 30 gave the trisodium salt of thallusin isomer 31.

Finally, the morphogenesis-inducing activities of the synthetic ( $\pm$ )-thallusin (23) and analog ( $\pm$ )-31 in comparison with natural thallusin (3), two natural product derivatives  $(32 \text{ and } 33)^1$  $(32 \text{ and } 33)^1$  and tricarboxylate 34 (the model compound used to develop the synthetic route, $5$  [Scheme](#page-3-0) [4\)](#page-3-0) were investigated in Monostroma oxyspermum.<sup>[16](#page-4-0)</sup> As



<span id="page-3-0"></span>

Scheme 4.

Table 1. Morphogenesis-inducing activity against M. oxyspermum after 7 days cultivation

Compound	$MEC$ (fmol/mL) <sup>a</sup>
Natural Thallusin (3)	3.8
Synthetic $(\pm)$ -Thallusin (23)	7.6
Natural product derivative 32	3900
Natural product derivative 33	15,600
31	1630
34	Inactive

<sup>a</sup> MEC was measured as the mean of the three independent tests.

shown in Table 1, natural thallusin (3) showed activity even at 3.8 fmol/mL,<sup>1</sup> and synthetic ( $\pm$ )-thallusin also showed comparable activity (at 7.6 fmol/mL). Interestingly, thallusin analog  $(\pm)$ -31 showed substantial activity (at 1630 fmol/mL) that was more potent than that of trimethyl ester 32 or the reduction product 33. Model compound 34 did not show any morphogenesis-inducing activity or growth inhibitory activity. These results suggest that the free carboxylic residues and picolinic acid moiety are indispensable for morphogenesis-inducing activity and that the diterpene skeleton also plays an important role.

Thus, we completed the total synthesis of  $(\pm)$ -thallusin based on the  $Hg(OTf)<sub>2</sub>-TMU$  complex induced biomimetic olefin cyclization and Suzuki coupling with pyridylboronic acid derivative as the key steps, and we developed a novel  $Hg(OTf)$ -catalyzed olefin isomerization leading to thermodynamically more stable isomer by using in toluene.

## Acknowledgment

This study was financially supported by a Grant-in-Aid from the Ministry of Education, Culture, Sports, Science, and Technology of the Japanese Government.

### Supplementary data

Supplementary data associated with this article can be found, in the online version, at [doi:10.1016/](http://dx.doi.org/10.1016/j.tetlet.2007.04.075) [j.tetlet.2007.04.075.](http://dx.doi.org/10.1016/j.tetlet.2007.04.075)

## References and notes

1. Matsuo, Y.; Imagawa, H.; Nishizawa, M.; Shizuri, Y. Science 2005, 307, 1598.

- 2. Gao, X.; Matsuo, Y.; Snider, B. B. Org. Lett. 2006, 8, 2123–2126.
- 3. (a) Nishizawa, M.; Takenaka, H.; Nishide, H.; Hayashi, Y. Tetrahedron Lett. 1983, 24, 2581–2584; (b) Nishizawa, M.; Morikuni, E.; Asoh, K.; Kan, Y.; Uenoyama, K.; Imagawa, H. Synlett 1995, 169–170; (c) Nishizawa, M.; Takenaka, H.; Hirotsu, K.; Higuchi, T.; Hayashi, Y. J. Am. Chem. Soc. 1984, 106, 4290-4291; (d) Nishizawa, M.; Takenaka, H.; Hayashi, Y. J. Am. Chem. Soc. 1985, 107, 522–523; (e) Nishizawa, M.; Takenaka, H.; Hayashi, Y. J. Org. Chem. 1986, 51, 806–813; (f) Nishizawa, M. Studies in Natural Product Chemistry. In Stereoselective Synthesis, Part A; Rahman, A., Ed.; Elsevier: Amsterdam, Holland, 1988; Vol. 1, pp 655–676; (g) Nishizawa, M. J. Syn. Org. Chem. Jpn. 1999, 57, 677–688; (h) Nishizawa, M.; Imagawa, H. J. Syn. Org. Chem. Jpn. 2006, 64, 744– 751.
- 4. (a) Nishizawa, M.; Skwarczynski, M.; Imagawa, H.; Sugihara, T. Chem. Lett. 2002, 12–13; (b) Nishizawa, M.; Yadav, V. K.; Skwarczynski, M.; Takao, H.; Imagawa, H.; Sugihara, T. Org. Lett. 2003, 5, 1609–1611; (c) Nishizawa, M.; Takao, H.; Yadav, V. K.; Imagawa, H.; Sugihara, T. Org. Lett. 2003, 5, 4563–4565; (d) Imagawa, H.; Kurisaki, T.; Nishizawa, M. Org. Lett. 2004, 6, 3679– 3681; (e) Imagawa, H.; Iyenaga, T.; Nishizawa, M. Org. Lett. 2005, 7, 451–453; (f) Imagawa, H.; Iyenaga, T.; Nishizawa, M. Synlett 2005, 703–705; (g) Imagawa, H.; Asai, Y.; Takano, H.; Hamagaki, H.; Nishizawa, M. Org. Lett. 2006, 8, 447–450; (h) Imagawa, H.; Fujikawa, Y.; Tsuchihiro, A.; Kinoshita, A.; Yoshinaga, T.; Takao, H.; Nishizawa, M. Synlett 2006, 639–641; (i) Imagawa, H.; Kotani, S.; Nishizawa, M. Synlett 2006, 642–644; (j) Imagawa, H.; Kinoshita, A.; Fukuyama, T.; Yamamoto, H.; Nishizawa, M. Tetrahedron Lett. 2006, 47, 4729–4731; (k) Yamamoto, H.; Nishiyama, M.; Imagawa, H.; Nishizawa, M. Tetrahedron Lett. 2006, 47, 8369–8373; (l) Kurisaki, T.; Naniwa, T.; Yamamoto, H.; Imagawa, H.; Nishizawa, M. Tetrahedron Lett. 2007, 48, 1871–1874.
- 5. (a) Miyaura, N.; Suzuki, A. Chem. Rev. 1995, 95, 2457– 2483; (b) Selles, P.; Mueller, U. Org. Lett. 2004, 6, 277– 279.
- 6. Tamao, K.; Ishida, N.; Kumada, M. J. Org. Chem. 1983, 48, 2120–2122.
- 7. Grieco, P. A.; Nishizawa, M.; Marinovic, N.; Ehmann, W. J. J. Am. Chem. Soc. 1976, 98, 7102–7104.
- 8. Camphausen, K.; Sproull, M.; Tantama, S.; Sankineni, S.; Scott, T.; Menard, C.; Coleman, C. N.; Brechbiel, M. W. Bioorg. Med. Chem. 2003, 11, 4287–4293.
- 9. Barrero, A. F.; Alvarez-Manzaneda, E. J.; Chahboun, R.; Cortes, M.; Armstrong, V. Tetrahedron 1999, 55, 15181– 15208.
- 10. Comins, D. L.; Dehghani, A. Tetrahedron Lett. 1992, 33, 6299–6302.
- 11. Voisin, A. S.; Bouillon, A.; Lancelot, J.-C.; Rault, S. Tetrahedron 2005, 61, 1417–1421.
- 12. Maruoka, K.; Murase, N.; Yamamoto, H. J. Org. Chem. 1993, 58, 2938–2939.
- 13. Bessard, Y.; Crettaz, R. Heterocycles 1999, 51, 2589-2602.
- 14. Duggan, A. J.; Adams, M. A.; Brynes, P. J.; Meinwald, J. Tetrahedron Lett. 1978, 4323–4326.
- 15. Crystal data for 27: A single crystal of 27 was obtained by the recrystallization from hexane/dichloromethane. The crystal was a colorless cube (dimensions  $0.20 \times 0.20 \times$ 0.20 mm) and was stable at room temperature. Measurements were performed with a Mac Science (Bruker Nonius) dip image plate diffractometer using graphitemonochromated Mo K<sub>a</sub> radiation ( $l = 0.71073$  Å). Cell parameters were as follows:  $a = 6.5980$  (3) Å,  $b = 23.6990$ (12) A<sup> $\zeta$ </sup> c = 13.8790 (8) A<sup> $\zeta$ </sup> b = 92.133 (2)°,  $V = 2168.7$

<span id="page-4-0"></span>(2)  $\mathring{A}^3$ , space group  $P2_1/c$ ,  $Z = 4$ ,  $D_x = 1.392$  Mg m<sup>-3</sup>,  $m = 0.080$  mm<sup>-1</sup>,  $T = 298$  K. A total of 3834 reflections were collected, of which 3770 were independent. The crystal structure was solved by the direct method with SIR-97. Refinement was performed by a full matrix least-<br>squares method on  $F^2$  with shell xlass-97. Anisotropic refinement was applied for all non-hydrogen atoms. All hydrogens could be located by difference Fourier methods. The final values of  $R$ , w $R$  factors and goodness-of-fit  $(S)$ were 0.0492, 0.1332, and 1.059, respectively. Crystallographic data for the structure of 27 were deposited with the Cambridge Crystallographic Data Centre as Supplementary Publication Nos. CCDC 635112. These data can be obtained free of charge via [www.ccdc.cam.ac.uk/](http://www.ccdc.cam.ac.uk/data_request/cif) [data\\_request/cif,](http://www.ccdc.cam.ac.uk/data_request/cif) by emailing data\_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12, Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

16. (a) Tatewaki, M.; Provasoli, L.; Pintner, J. J. Phycol. 1983, 19, 409–416; (b) Provasoli, L. Proc. Int. Seaweed Symp. 1963, 4, 1–17; (c) Matsuo, Y.; Suzuki, M.; Kasai, H.; Shizuri, Y.; Harayama, S. Microbiol 2003, 5, 25–35.